How to use... a blood film

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ABSTRACT

The diagnostic relevance of the blood film cannot be underestimated in the assessment of children with suspected primary or secondary haematological conditions. The blood film not only serves as a diagnostic tool but also allows for screening, monitoring of disease progression and therapeutic response in children with a variety of haematological conditions. This article outlines the appearance of normal paediatric and neonatal blood films. The technical aspects involved in preparing a blood film are discussed. Consideration is given to the indications for preparing a blood film and some of the limitations of blood films. Finally, attempts are made to highlight the role of the blood film in the diagnosis of some common paediatric and neonatal conditions.

INTRODUCTION

The blood film is the preparation of blood on a slide for microscopic analysis of peripheral blood cells. The development of automated blood cell analysers has led to a reduction in the number of blood samples requiring a blood film. However, the diagnostic relevance of the blood film cannot be underestimated in the assessment of children with suspected primary or secondary haematological conditions. The blood film not only serves as a diagnostic tool but also allows for screening, monitoring of disease progression and therapeutic response in children with a variety of haematological conditions including leukaemia. haemolytic uraemic syndrome and bone marrow failure syndromes such as Fanconi anaemia.1 We believe it is important for paediatricians to have an understanding of blood film interpretation in order to correctly apply results to their clinical practice. This paper highlights the indications for preparing a blood film and an overview of the possible findings.

PHYSIOLOGICAL BACKGROUND/THE NORMAL BLOOD FILM

Haemopoiesis mainly occurs in the liver of the fetus. After birth and throughout life, haemopoiesis occurs in the bone marrow. In infants and the growing child, this occurs in all bones of the skeleton.²

An understanding of the normal cytology is essential in order to allow for identification of abnormalities on the blood film. Red cells are the most numerous cell type encountered in the blood film. In the paediatric film, normal red cells are the size of the lymphocyte nucleus with a diameter of 7-9 µm and a mean corpuscular volume (MCV) of 75-90 fL. They should be round in shape with a smooth contour appearing as a biconcave disc. Approximately, one-third of the cell should have a central pallor.³ The neonatal blood film differs from the paediatric blood film. It is not uncommon to see burr cells (echinocytes), occasional nucleated red blood cells (RBC), target cells, fragmented red cells and some spherocytes. Neonates typically have an elevated MCV and red cells are therefore macrocytic.4

White blood cells can be divided into the myeloid/monocytic cells (neutrophils, eosinophils, basophils and monocytes) and lymphocytes. Segmented neutrophils are the predominant white cells in the peripheral blood. The total white cell count and the neutrophil, monocyte and lymphocyte counts are often much higher in the neonate than the older child. In addition, it is important to remember that the automated lymphocyte count may be falsely elevated due to the presence of nucleated red blood cells.

Platelets are small, non-nucleated cells. They normally measure $1.5\text{--}3\,\mu m$ in diameter. They are derived from the cytoplasmic fragments of megakaryocytes. 3 4

TECHNOLOGICAL BACKGROUND

A venous or capillary blood sample should be mixed with a suitable anticoagulant such as EDTA in a sample bottle and transported to the lab without undue delay. It



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Figure 1 Example of both a stained and unstained blood film.

is very helpful to the laboratory staff to include clinical information on the request form.

The blood film is prepared by skilled laboratory staff or by an automated blood film machine. The sample should be well mixed. A small drop of blood is dropped onto a clean glass slide. A smear is then made across the slide using the wedge technique. The slide is then stained before examination. Most laboratories use a variant of the Romanosky stain. A completed blood film is shown in figure 1.

The blood film should then be examined by microscopy with a systematic approach by staff trained in haematology morphology. The morphology of the blood cells on a blood film is best assessed by looking at the three cell groups-red cells, white cells and platelets. The size, shape, colour of the red cells and the presence of red cell inclusions should be assessed. White cell morphology should be assessed and a differential white cell count performed. Platelet count and size should be reviewed for any abnormalities. In addition, the blood film may also reveal other abnormal cells, for example, leukaemic blast cells. Infections which may be evident from blood film analysis include malaria, borrelia, filarial and candida albicans. It is essential that the blood film is always interpreted alongside the patient's clinical details.

INDICATIONS/LIMITATIONS

There are two main reasons why a blood film may be created.

- 1. Request of the clinician based on clinical concerns.⁷
- 2. Laboratory criteria following an automated complete blood count and differential white cell count as suggested by the International Consensus Group for Haematology Review in 2005.8

There are some limitations associated with blood films.

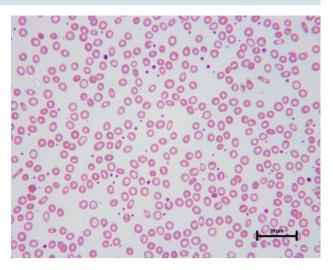


Figure 2 Blood film showing hypochromic, microcytic red cells.

- 1. Delays in transporting samples to the laboratory and exposure to extremes of temperature can cause artefactual changes.
- Clotted samples cannot be accurately used and often need to be repeated.
- 3. Poor technique leads to the creation of poor quality slides which can be difficult to interpret.

CLINICAL OUESTIONS

This section uses four clinical questions to discuss the use of blood films in common and important paediatric conditions.

Can iron deficiency anaemia be diagnosed from a blood film?

The presence of hypochromic, microcytic red cells most often suggests iron deficiency anaemia (IDA) (figure 2). However, this picture may also be present in patients with thalassaemia. It is essential to differentiate between these conditions to avoid inappropriate iron therapy in patients with thalassaemia. Standard tests to differentiate between IDA and thalassaemia include iron studies, haemoglobin electrophoresis and molecular testing for haemoglobin variants.

Beta thalassaemia major and haemoglobin H disease may be distinguished from IDA by blood film morphology due to the increased presence of target cells, irregularly contracted cells and basophilic stippling. However, in the thalassaemia trait syndromes, the differences are much less marked. It is therefore more useful to examine the RBC indices in these cases to distinguish them from IDA. In thalassaemia, the mean corpuscular haemoglobin concentration (MCHC) is often normal. This contrasts to IDA where the MCHC is often low. In thalassaemia, the haemoglobin level tends to be higher relative to the MCV than would be expected in IDA. Finally, although there is a mild microcytic anaemia in both conditions, the red cell distribution width is usually only elevated in IDA.

By way of example, consider these two sets of blood results: (table 1)

High

The state of a patient of a pat					
	Thalassaemia		Iron deficiency	Iron deficiency anaemia	
Haemoglobin	101 g/L	Low	91 g/L	Low	
Haematocrit	0.31	Low	0.3	Low	
Platelet count	233×10 ⁹ /L	Normal	453×10 ⁹ /L	High	
Red blood cells	5.2×10 ¹² /L	Normal	4.67×10 ¹² /L	Normal	
Mean corpuscular volume	58 fL	Low	64.2 fl	Low	
Mean corpuscular haemoglobin concentration	327 g/L	Normal	303 g/L	Low	
Mean corpuscular haemoglobin	19 pg	Low	19.5 pg	Low	

Full blood picture of a patient with thalassaemia and a patient with iron deficiency anaemia

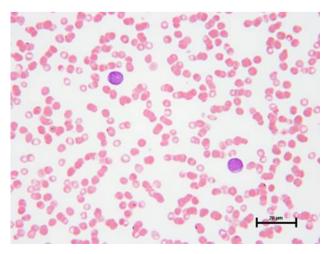
15.4%

Does a normal blood film exclude leukaemia?

Red cell distribution width-coefficient of variation

Leukaemia is the most common childhood malignancy accounting for approximately 30% of cases. Clinical presentation can be highly variable depending on the child's age and the extent of leukaemic infiltration of the bone marrow and other sites. A British Medical Journal review on the diagnosis and management of acute leukaemia in children cited the full blood picture and blood film as the most useful initial investigations when there is a clinical suspicion of leukaemia. 10 The full blood picture will often reveal pancytopenia due to bone marrow infiltration. Blast cells may elevate the white cell count despite neutropaenia. The presence of blast cells in the blood film is highly indicative of leukaemia. (figure 3) Blast cells are immature cells with open nuclear chromatin. They vary in size from the size of small mature lymphocytes to large cells. Many have a high nucleus:cytoplasmic ratio. The nuclei may be irregular in shape. Other features may include Auer rods and cytoplasmic granulation or vacuolisation. They are best visualised at the lateral edges of the film.

However, a normal full blood picture and blood film does not exclude leukaemia. Patients who do not have bone marrow suppression may have a normal full blood picture. In addition, the peripheral blood film may be normal if blast cells are confined to the bone marrow. If leukaemia is clinically suspected, further investigations which may be warranted include:



Blood film showing blast cells in patient with leukaemia.

bone marrow aspiration and biopsy for definitive

17.5%

imaging to assess the extent of disease;

Normal

- lumbar puncture in cases of suspected central nervous system infiltration;
- cytogenetic analysis and immunophenotyping.

Does a normal blood film in a newborn with trisomy 21 exclude clinically significant transient abnormal myelopoeisis (TAM)?

Children with trisomy 21 have an increased risk of leukaemia. In addition, they are at risk of TAM, a haematological disorder found exclusively in neonates with trisomy 21. TAM is thought to be a disorder of fetal haemopoeisis. It appears to be caused by a single GATA1 mutation in children with constitutive trisomy 21.¹¹ Estimates suggest that 10% of neonates with trisomy 21 will have TAM. 11 Approximately, 25% of neonates with TAM are asymptomatic. In those who are symptomatic, the clinical features are variable and include:

- iaundice:
- bleeding problems;
- respiratory distress;
- liver failure.

As TAM is a disorder of haemopoeisis, the blood film of affected patients will be abnormal. Common findings include elevated numbers of circulating, immature myeloid cells, including basophilic blasts, nucleated

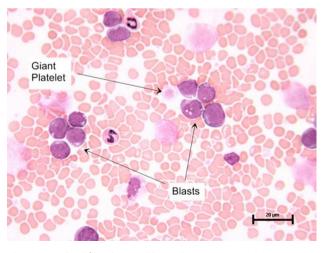


Figure 4 Blood film showing blast cells in neonate with transient abnormal myelopoeisis.

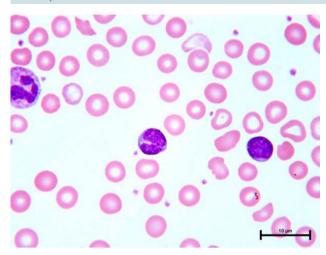


Figure 5 Blood film showing small lymphocytes with hyperchromatic, cleaved nuclei.

red cells, megakaryocyte fragments and thrombocytosis or thrombocytopaenia¹² (figure 4). However, it is not uncommon for the blood film to be abnormal in neonates with trisomy 21. A recent prospective analysis of 200 neonates with trisomy 21 showed that 195 of these neonates (97.5%) had circulating blasts. 11 In the majority of neonates, TAM will resolve spontaneously without any intervention. Those with severe symptoms may require treatment with low-dose cytarabine chemotherapy. Approximately, 20% will progress to develop acute megakaryoblastic leukaemia before 4 years of age.

Is a blood film of use when a diagnosis of pertussis is suspected?

Pertussis is an acute bacterial infection caused by Bordetella pertussis. Studies have shown that up to 50% of children with pertussis have an absolute lymphocytosis. Pertussis lymphocytosis is caused by pertussis toxin released by B. pertussis but the exact mechanisms by which this occurs remain unclear. 13 On morphological examination of the mature lymphocytes, they appear small with hyperchromatic, cleaved nuclei (figure 5).14 Therefore, the blood film may prove a useful diagnostic aid in patients with unexplained lymphocytosis.

CONCLUSION

The blood film is a quick and important investigative tool. An understanding of the appearance of a normal paediatric and neonatal blood film is crucial to accurate interpretation. The blood film has a role in both the diagnosis and monitoring of a number of paediatric haematological conditions. A systematic approach to examination of the blood film is essential. It is always important to interpret the result in correlation with the clinical picture.

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Competing interests None declared.

Clinical bottom line

- Although hypochromic microcytic anaemia often represents iron deficiency anaemia in children, careful analysis of all red cell values reported on the complete blood count and examination of the blood film will help to distinguish between iron deficiency anaemia and thalassaemia.
- The presence of blast cells in the blood film is highly indicative of leukaemia. However, a normal full blood picture and blood film does not exclude leukaemia.
- The blood film of neonate with trisomy 21 and clinical signs of transient abnormal myelopoeisis will be abnormal and may display elevated numbers of circulating, immature myeloid cells, including basophilic blasts, nucleated red cells, megakaryocyte fragments and thrombocytosis or thrombocytopaenia.
- Characteristic changes in lymphocytes on the blood films in patients with a lymphocytosis may help with diagnosing Bordetella pertussis.

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REFERENCES

- 1 Adewoyin AS, Nwogoh B. Peripheral blood film a review. Ann Ib Postgrad Med 2014;12:71-9.
- 2 Gordon-Smith T. Haemopoiesis the formation of blood cells. Medicine 2009;37:129-32.
- 3 Hays T, Jamieson B. Atlas of paediatric peripheral blood smears. 1st edn: Abbott Laboratories, 2008.
- 4 Bain BJ, Bates I, Laffan MA. Dacie and Lewis practical haematology: Elsevier Health Sciences, 2016.
- Bain BJ. Blood cells: a practical guide: John Wiley & Sons, 2015.
- 6 Tkachuk DC, Hirschmann JV. Wintrobe atlas of clinical haematology: Lippincott Williams & Wilkins, 2007.
- Bain BJ. Diagnosis from the blood smear. N Engl J Med 2005;353:498-507.
- 8 Barnes PW, McFadden SL, Machin SJ, et al. The international consensus group for hematology review: suggested criteria for action following automated CBC and WBC differential analysis. Lab Hematol 2005;11:83-90.
- Trivedi DP, Shah HA. Discriminant functions in distinguishing beta-thalassemia trait and iron deficiency anemia: the value of the RDW-SD. Internet J Hematol 2011;7:1-13.
- Chris M, Georgina H, Clarke Rachel T. Acute leukaemia in children: diagnosis and management. BMJ 2009;338:b2285.
- 11 Bombery M, Vergilio JA. Transient abnormal myelopoiesis in neonates: GATA get the diagnosis. Arch Pathol Lab Med 2014;138:1302-6.
- 12 Roberts I, Alford K, Hall G, et al. GATA1-mutant clones are frequent and often unsuspected in babies with Down syndrome: identification of a population at risk of leukemia. Blood 2013;122:3908-17.
- 13 Carbonetti NH. Pertussis leukocytosis: mechanisms, clinical relevance and treatment. Pathog Dis 2016;74:ftw087.
- 14 Pandey S, Cetin N. Peripheral smear clues for Bordetella pertussis. Blood 2013;122:4012.